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Review

Adaptations to subordinate status in female marmoset monkeys

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Abstract

In singular cooperatively breeding mammals, social status is a key determinant of female reproductive success. Usually only one dominant female breeds in a social group. In the common marmoset (*Callithrix jacchus*), a cooperatively breeding primate, adaptations to nonbreeding subordinate status appear to parallel those found in social insect societies which demonstrate similar skew to the marmoset in female reproductive success. Female marmosets exhibit rank-related polyethism in behavior, reproductive neuroendocrinology and non-reproductive physiology, and subordinates participate in alloparental care and territory defense. Olfactory, visual and behavioral cues from dominant female marmosets, therefore, appear to have developed specific neural and neuroendocrine adaptations to subordinate social status analogous to those found in social insects such as the lower wasps, bees and termites. Such parallel developments probably reflect the outcome of repeated convergent evolutionary attempts at adapting to environmental conditions not readily conducive to dispersal and independent breeding. © 1998 Elsevier Science Inc. All rights reserved.

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1. Introduction

Food availability, climate and the social environment represent the major ultimate factors regulating reproductive success in female mammals [18]. Proximate cues predictive of favorable food availability and climate serve to activate changes in reproductive physiology, behavior and metabolism, culminating in the optimal timing of courtship, mating, parturition and weaning of offspring [59]. The social environment, however, has a more complex impact on reproductive success and provides a plethora of proximate cues. Social factors regulate the timing of reproductive success in two distinct ways which are not mutually exclusive. First, social factors can act to modulate the timing of reproduction, when food availability and climate are either approaching or have already reached optimal conditions. For example, olfactory, visual and behavioral cues from rams advance and synchronize estrus in ewes. Increased concentrations of luteinizing hormone (LH) are measurable in the ewes' circulation within minutes of exposure to male cues [62]. However, this stimulatory effect of males occurs only just prior to or at the beginning of the breeding season [23,87].

Second, in addition to modulating the timing of reproduction, social factors can serve as constraints on reproductive success over and above those imposed by food availability and climate. Thus, social factors may effectively 'gate' the relevance of cues related to the physical environment to times when the social environment is suitable for successful reproduction, e.g. when

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high rank or dominant status is attained. In this regard, low social rank or subordinate status has been widely shown to inhibit reproductive function in females in a variety of species [17,62,69] with varying intensity and effectiveness of inhibition [26,34,101].

Many mammalian species are characterized as competitive breeders, in which most or all females in each social group breed. In such species, however, subordinate females may experience lower reproductive success than higher ranking females [61]. Mechanisms by which dominant females may secure their reproductive advantages over subordinate females include: (1) harassmentinduced disruption of subordinates' sexual behavior (e.g. stumptail macaque, *Macaca arctoides* [32]); (2) harassment-induced stress in subordinates, resulting in inhibited ovulation (e.g. cynomolgus monkeys, Macaca fascicularis [53]) or failed implantation following conception (e.g. Syrian hamsters, Mesocricetus auratus [50]); (3) harassment-related infant loss experienced by subordinates (e.g. yellow baboons, Papio cynocephalus [70]); and (4) exclusion of subordinate females from resources essential for successful reproduction, such as food (e.g. red deer, Cervas elephas [22]).

In contrast to the graded reproductive suppression found in most competitive breeders, some of the most extreme mammalian examples of dominance-determined female reproductive success are found in cooperatively breeding species. Within social groups of cooperatively breeding species, members other than the genetic parents play crucial roles in successfully rearing offspring. In singular cooperative breeders, reproduction is limited to a single female in each group [29,52]. Reproductive failure in subordinate females in these species may not be dependent upon generalized harassment or resource exclusion [10]. Instead, specialized neuroendocrine and behavioral responses by females to subordinate status may directly inhibit sexual behavior (e.g. golden lion tamarins, *Leontopithecus rosalia*; [40]), ovulation (e.g. common marmosets, Callithrix jacchus; [9]) and implantation (e.g. white-footed mice, Peromyscus leucopus; [41]). Olfactory, visual and behavioral cues from dominant females have been implicated in the activation or maintenance of specific inhibitory reproductive mechanisms in such subordinate females, indicating a well-developed sensitivity and specialized neuroendocrine response to proximate cues signifying subordinate status [10].

Singular cooperatively breeding species are found throughout the avian and mammalian classes and are characterized by delayed dispersal of offspring from the natal group, reproductive suppression and care for nondescendant offspring [91]. These specialized characteristics resemble those of eusocial insects, in which colonies contain overlapping generations, clear divisions of labor and of reproduction occur, and nonbreeding members cooperate in rearing offspring and in protecting and servicing the colony [65,103].

Such similarities in definitional characteristics between cooperative breeders and eusocial insects have prompted Sherman and colleagues [86] to reconsider eusociality as a continuum across which societies of cooperatively breeding vertebrates and invertebrates can be arranged depending upon the degree of skew shown in female reproductive success within social groups (Fig. 1). Such an inclusive concept emphasizes the behavioral similarities between cooperatively breeding vertebrates and invertebrates, encompasses all forms of alloparental care, and removes the traditional distinctions between cooperative breeding and eusociality. Lacey and Sherman [57] have proposed a single integrated hypothesis for the evolution of singular cooperative breeding in both vertebrates and invertebrates. In slightly modified form, their scenario suggests that for cooperative breeding to evolve: (1) ecological constraints on dispersal and on independent breeding became so onerous that coloniality or social grouping was favored; and (2) alterations in the patterns of intra-group relatedness (increased kinship), reproductive biology (development of alloparental care and reproductive suppression among subordinates) and phenotype (somatic and physiological adaptations to social status) further promoted cooperation within such colonies or social groups. In other words, comparable environmental constraints on female reproduction across vertebrate and invertebrate cooperatively breeding species may result in the convergent evolution of similar proximate mechanisms. To date, the concept of a continuum of eusociality has been addressed mainly at the ultimate level of causation. Here, we aim to take the argument to the proximate level and to provide evidence to stimulate further detailed examination.

Our research has focused on proximate mechanisms mediating social regulation of female reproductive success in the common marmoset, a cooperatively breeding anthropoid primate. Our laboratory findings suggest that females have specialized adaptations to subordinate status in terms of behavior, reproductive neuroendocrinology, hypothalamic-pituitary-adrenal function and glucose homeodynamics. In this review, we will discuss how behavioral and physiological consequences of subordinate status in female marmosets appear to reflect a stable alternative to dominant status rather than a state of generalized stress imposed by the dominant female and endured by subordinates to their physiological detriment. We will explore how the social environment provides the predominant cue(s) timing female reproductive effort in marmosets, and we will consider anovulation as just one of many specialized adaptations to subordinate status in this cooperatively breeding species.

Based on the invertebrate literature dealing with cooperative breeding, the adaptations made by female marmosets to subordinate status can be considered as

Eusociality Continuum



Fig. 1. Diagrammatic representation of predicted locations of selected cooperatively breeding vertebrate and invertebrate species along the eusociality continuum [86], arranged with regard to an index of the skew in estimated lifetime reproductive success of females in a social group [55,86]. Skew equals '0' when female lifetime success is equal among group members and reaches a value of '1' when reproduction is limited to one female in a group. As skew may vary among conspecific groups or populations, species having approximately similar degrees of skew are grouped together. The figure is modified from Lacey and Sherman [57].

examples of polyethism, even if they are reversible with a change in social status [16]. Consequently, we aim to review the degree and nature of polyethism found in female marmosets, as a first step in ascertaining the commonality of proximate mechanisms shaping the specialized adaptations of cooperatively breeding species across vertebrate and invertebrate classes. Such comparative examination of the proximate mechanisms regulating female reproductive success may provide novel insights into the neural mechanisms mediating environmental control of neuroendocrine function.

2. Common marmosets

Along with the other members of the family Callitrichidae (marmosets and tamarins), the common marmoset is a small New World primate exhibiting a singular cooperative breeding strategy. In the wild, common marmosets occupy successional forest, forest edge habitat and savanna gallery forest in northeastern Brazil [75]. A characteristic feature of this species is its exploitation of exudate feeding supplemented by opportunistic feeding on fruits, insects and small vertebrates, which has enabled its habitat expansion despite the extensive deforestation of Atlantic coastal forests.

2.1. Social groups, reproductive division of labor and cooperative behavior

Free-living groups of common marmosets contain 3–15 individuals, usually including 2–4 adults of each sex [28,38]. Groups appear to consist mainly of extended families and possibly unrelated immigrants [39,64]. Offspring remain within their natal group into adulthood, and all group members contribute to infant care, including post-weaning provisioning of infants [40,97].

Both field [49,92] and laboratory [1,30,73,77] studies of common marmosets typically report that only a single, dominant female breeds in each social group but see [28]. Lack of 'helpers' to raise offspring, lack of dispersal opportunities, infanticide by the dominant female, and inbreeding avoidance may contribute to the ineffective breeding of subordinate females in free-living marmoset groups [12,27,40,71,77]. Subordinate females can sometimes breed successfully in free-living [27,71] and captive [11,74] groups, but these occurrences may depend on the subordinate's parturition occurring more than 1 month before or after the dominant female gives birth, increased food availability, large group numbers (increased numbers of helpers) and high population density (increased need for retention of subordinates for territorial defense) [40].

Callitrichids may migrate into established groups [37,83]. The successful founding of a new group by a single breeding pair appears unlikely because of the relatively high costs of gestation and infant care [40,97] and the minimum quotient of 'helpers' needed to successfully rear the infants of a single breeding female [95]. Thus, cooperation between animals may be required for the successful founding of new marmoset groups, as well as for their maintenance.

2.2. Standardized captive social groups

To investigate the proximate behavioral, neuroendocrinological, physiological and sensory regulation of reproductive success in female marmosets, we have established a total of 75 mixed-sex social groups comprising three to seven unrelated adults or postpubertal animals, over a 12-year period, at the Wisconsin Regional Primate Research Center in Madison, WI [79] and at the Institute of Zoology in London, UK [8]. Social groups remain together for 2 months to over 3 years. The social structure of our standardized captive social groups is typical of that for common marmosets. Linear or despotic hierarchies are usually quantifiable within 3 days of group formation, from the directionality of agonism displayed [1,76]. Dominant females are usually the only reproductively active females.

Endocrine monitoring of ovarian activity is accomplished by assessing plasma progesterone concentrations in twice weekly blood samples, which identifies the peri-ovulatory period (day of ovulation +1 day), phases of the ovarian cycle and anovulatory periods [8,45,79]. Stable group composition is achieved by contraceptive management of females with injections of a prostaglandin F2 α analogue [94]. Routine handling of the animals and blood collection procedures do not result in obvious physiological disturbances or in the disruption of the normal diurnal rhythm of plasma cortisol concentrations [79]. Plasma concentrations of cortisol, dehydroepiandrosterone (DHEA), adrenocorticotropic hormone (ACTH) and fasting glucose are determined from blood samples taken within 3 min of the investigator entering an animal's cage.

3. Behavioral polyethism

Dominant (breeding) and subordinate (non-breeding) females show clear behavioral differences. Within groups in the wild [28,48,92] and in the laboratory [3,13,30,49,73] subordinate females engage in little or no sexual activity. Outside their own group, however, subordinates may readily engage in sexual interactions [3,13,48,49], suggesting that inhibition of intra-group sexual behavior might depend on the presence of a dominant female groupmate and/or a familiarity or incest taboo [3,13,82]. Dominant females also engage in more affiliative interactions with the dominant male than do subordinate females in captive groups [1].

In free-living and captive groups, subordinates participate more in intergroup displays than dominants ([56]; C. Lazaro-Perea and C.T. Snowdon, personal communication). These findings may indicate a significant contribution by subordinates to territorial defense and rank-related divisions of labor.

4. Reproductive polyethism

4.1. Ovarian and pituitary function

Ovarian size, content and function differ dramatically between dominant and subordinate female marmosets. The average ovarian volume of subordinate females $(609 + 145 \text{ mm}^3; \text{ mean} + \text{S.E.M.}; n = 5)$ is approximately half that found in dominant females in the early to mid-follicular phase of the ovarian cycle $(1217 \pm 163 \text{ mm}^3; n = 8; P < 0.01 \text{ vs. subordinate fe-}$ males; Student's *t*-test). The absence of corpora lutea and corpora albicantia in ovaries of subordinate females suggests a lack of recent ovulatory function [9,100]. Ovarian follicle content also differs dramatically between dominant and subordinate females. Ovarian dissection revealed only 80-120 small (≤ 1.0 mm), antral follicles from each subordinate, whereas approximately twice that number of antral follicles were recovered from the ovaries of each dominant [43,44,46,47]. In all, 12% of follicles from the ovaries of dominant females exceeded 1.0 mm in diameter and 3% were preovulatory. Clearly, ovarian function in subordinate female marmosets was severely impaired.

Impaired ovarian function in subordinates was further confirmed by endocrine monitoring. Whereas dominant females ovulate regularly in captive social groups, subordinate females usually exhibit hypogonadotropic anovulation (Fig. 2) [8]. Serial blood sampling every 15 min for 4 h by repeated venipuncture [7] reveals a stark contrast between the low, nonepisodic plasma LH concentrations of subordinates and the higher, episodic plasma LH concentrations of dominant females in the mid-follicular phase of the ovarian cycle (Fig. 3a). Hypogonadotropism in subordinates is rapidly reversed by a change in social environment. Removal of an anovulatory subordinate from her dominant female results in an elevation of LH levels within 1-4 days [6,8], leading to the induction of ovulatory function almost within the length of a normal follicular phase [15]. Hypogonadotropic anovulation is readily re-engaged when ovulatory female marmosets are returned to subordinate status in their original group or in a new group [8,14].

These findings highlight the remarkably labile pituitary and ovarian responses of female marmosets to changes in social status. This dynamic reproductive neuroendocrine system may be an adaptation to cope with changing environments, varying group composition and an opportunistic existence ([64,75,84]; M.E. Yamamoto and D. Santee, personal communication). Subordinate females also show variability in the degree and duration of hypogonadotropism and ovarian inhibition depending on their subordinate rank, age and group composition [6,77,80]. For example, rank 2 subordinates exhibit less reliable hypogonadotropism and anovulation than rank 3 or 4 subordinates [6], and they may rapidly engage in sexual behavior and commence regular ovulatory cycles following the replacement of the breeding male by an unfamiliar (or unrelated) male



Days from Prostaglandin Analogue Administration

Fig. 2. Plasma concentrations of progesterone (solid circles) and immunoreactive LH (open circles) in (a) a typical dominant female marmoset during an ovarian cycle, and (b) an anovulatory subordinate female following i.m. administration of a prostaglandin F_2 analogue [94]. The elevated LH levels in the dominant female following the prostaglandin analogue treatment and approximately 20 days into the succeeding luteal phase, probably reflect cross-reactivity of the LH assay antibody with circulating levels of chorionic gonadotropin (CG) secreted from the placenta during early pregnancy.

[80,82]. Such a scenario might precipitate the replacement of the original breeding female by a previously subordinate female or the fissioning of the original group into new breeding groups.

4.2. Reproductive neuroendocrine inhibition

There is no evidence of generalized stress in anovulatory subordinate female marmosets. Body weight and diurnal rhythms are unaltered, and there are no elevations in the circulating concentrations of cortisol and prolactin [10,76,79]. Instead, specific inhibitory neuroendocrine mechanisms appear engaged in the maintenance of anovulation in subordinate females. Since pulsatile circulating LH levels usually correspond reliably with pulsatile release of gonadotropin-releasing hormone (GnRH) from the hypothalamus in other mammalian species (e.g. sheep: [58]; rhesus monkey, M. mulatta: [99]), the low, non-episodic pattern of circulating LH levels found in anovulatory subordinate female marmosets were initially interpreted as providing indirect evidence for the disruption or inhibition of hypothalamic GnRH release as the cause of the hypogonadotropism. We used two different approaches to test this hypothesis.

Our first approach involved treatment of anovulatory subordinates with exogenous GnRH. Subordinates received approximately hourly subcutaneous infusions of 1 or 2 μ g GnRH through an indwelling cannula, which was attached to a battery-powered syringe pump housed in a lightweight backpack [5,10]. GnRH treatment induced rapid elevations in circulating LH levels and ovulation within 2 weeks [10]. Two of six GnRHtreated subordinates even became pregnant during the treatment. On termination of GnRH treatment after 1-3 months, the subordinates quickly reverted to their hypogonadotropic state. Rapid induction of ovulatory function in subordinate females by GnRH treatment and the subsequent cessation of ovulatory function when GnRH treatment stopped, are similar to effects on ovulatory function obtained by simply manipulating female social status [8]. These results appeared to implicate pituitary hypogonadotropism in the mechanism of anovulation, and disrupted or inhibited hypothalamic GnRH release in the mechanism of hypogonadotropism.

To directly determine whether disrupted or inhibited release of endogenous hypothalamic GnRH accompanies hypogonadotropism in subordinate females, we measured dynamic GnRH release from the hypothalamus of conscious animals [10,81]. As it is not possible to accurately measure dynamic patterns of hypothalamic GnRH release from peripheral plasma concentrations [21,98], we employed a push-pull perfusion system to measure GnRH concentrations in 10-min perfusate fractions of artificial cerebrospinal fluid collected from



Fig. 3. (Continued)

6

20

0

the pituitary stalk-median eminence (S-ME) of marmosets. Throughout each perfusion, marmosets were restrained in a soft harness opposite their familiar groupmates [10,81].

In contrast to plasma LH concentrations, hypothalamic GnRH concentrations did not differ between dominant and subordinate females. Extra-cellular release of GnRH from the S-ME of anovulatory subordinates

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was pulsatile in nature (Fig. 3b), and GnRH was found in similar concentrations to those of dominant females in the mid-follicular phase of the ovarian cycle (Fig. 3b,c) [10,81]. Discrepancies between hypothalamic GnRH release and circulating LH levels have also been recently found in female rats during lactational anestrous [19]. Both of these findings, however, are unconventional. Previously, studies had demonstrated that hypogonadotropism is accompanied by, and is indeed caused by reduced hypothalamic GnRH release, such as in prepubertal female rhesus monkeys [102] and in seasonally anestrous ewes [54]. Certainly in the marmoset, the apparent qualitative differences between hypothalamic GnRH and pituitary LH activity in subordinate females leaves open the possibilities that subtle alterations in hypothalamic GnRH release may be reducing pituitary gonadotropic responsiveness to GnRH or that the primary site for gonadotropic failure may reside at the level of the pituitary. Factors such as reduced pituitary gonadotropic responsiveness to GnRH [8], insufficient endocrine priming of pituitary gonadotrophs [33,104,105], or a combination of factors leading to reduced pituitary sensitivity to GnRH [20], may be responsible for the hypogonadotropic condition of subordinates. Mediation of hypogonadotropism in subordinates by a subtle combination of neuroendocrine and endocrine disruption, rather than by a dramatic change in hypothalamic GnRH release, would be consistent with the extremely rapid changes in LH secretion seen when the social status of female marmosets is systematically manipulated [6,8]. Further elucidation of this possibility must await the outcome of comprehensive quantitative analyses of endogenous GnRH and LH secretion dynamics in dominant and subordinate female marmosets.

Why, then, had exogenous GnRH therapy been so effective in reversing the hypogonadotropism of subordinate females, if pituitary inadequacy plays a major role in its mechanism? The answer may lie in the relatively large doses of exogenous GnRH given during infusion treatments. Each subordinate female marmoset received 1 or 2 μ g GnRH intravenously or subcutaneously, infused at approximately hourly intervals, over a period of several weeks or months [5,10]. These GnRH amounts are well in excess of the 200 ng GnRH given intramuscularly to induce LH release from the pituitary of subordinate females [8] and are similarly in excess of endogenous peak levels of GnRH in hypothalamic push-pull perfusates (Fig. 3b,c). Such supraphysiological episodic GnRH stimulation of pituitary gonadotrophs may have overridden any endogenous pituitary hyporesponsiveness and sufficiently engaged pituitary gonadotropin function to induce ovarian follicular growth, ovulation and normal luteal function [4,10].

Evidence of specific ovarian hormone-dependent and ovarian hormone-independent mechanisms underlying hypogonadotropism has been obtained from manipulation of circulating estradiol levels in female marmosets [2]. As evidence for the former type of mechanism, subordinate females have demonstrated enhanced sensitivity to the inhibitory influences of estradiol feedback on LH secretion. In the case of the latter, experiments employing the opiate μ - and ε -receptor antagonist, naloxone, appear to implicate the inhibitory endogenous opioid peptides in the suppression of LH in ovariectomized, but not intact, subordinate females [5]. Endogenous opioid peptides have also been implicated in estradiol-independent mechanisms of steroid hormone negative feedback regulation of LH secretion in other primates [36]. The inhibitory neuroendocrine mechanisms operating in subordinate female marmosets may, thus, encompass a generalized enhancement of both estradiol-dependent and estradiol-independent negative feedback on LH secretion at hypothalamic and, possibly, pituitary levels. Similar dichotomies in inhibitory neuroendocrine regulation of LH secretion have been found in lactating rats [19,60,88] and in seasonally anestrous ewes [54,66]. Nevertheless, the precise nature of the inhibitory components involved in the neuroendocrine mechanism(s) inducing and maintaining such naturally-occuring hypogonadotropism still await elucidation.

Fig. 3. (a) Typical individual plasma bioactive LH concentrations in two mid-follicular phase dominant female marmosets (271-313W) and two anovulatory subordinate females (315-313W). The plasma samples were obtained without anesthesia from repeated femoral venipuncture every 15 min over four consecutive hours. In contrast to LH values in subordinate females, those in the dominants were higher in value and exhibited an episodic pattern. Note that the LH data from female 313W clearly differentiate when she was dominant in a group (high plasma values and episodic pattern), and then later when she was subordinate (low plasma values and nonepisodic pattern). A quantitative assessment of endogenous LH pulsatility was not performed because of the relatively long inter-sample interval between plasma LH determinations (15 min) relative to the total sample interval (4 h) and a putative pulse frequency of approximately 40–60 min. (b) Typical individual immunoactive GnRH concentrations in 10-min fractions of hypothalamic push-pull perfusate obtained without anesthesia from a female marmoset (CJ0086) while holding (1) dominant status in a social group (during the mid-follicular phase of the ovarian cycle), and later (2) subordinate status in another group (anovulatory). * Denotes GnRH pulses identified by the computer algorithm PULSAR [98] under both social conditions. Modified from Abbott et al. [10]. (c) Antilog of the transformed mean (+95% confidence limit) of GnRH baseline, interpulse intervals and peak concentration in six females in the midfollicular phase of the ovarian cycle (cycling, solid bars) and in six anovulatory subordinate females (hatched bars). Probability values were derived from Student's *t*-tests.



Fig. 4. Diagrammatic representation of the role of olfactory, visual and behavioral cues from dominant female marmosets in maintaining the inhibitory neuroendocrine mechanisms regulating hypogonadotropic anovulation in subordinate females.

5. Proximate cues regulating the social inhibition of female reproductive neuroendocrinology

Olfactory, visual and behavioral cues from dominant female marmosets all play a role in maintaining inhibition of ovulation in subordinate females [14]. Marmosets are one of the few primates which possess a fully functional accessory olfactory system and vomeronasal organ in addition to the main olfactory system [51,64,96] and specialized sternal, suprapubic and anogenital scent glands [31]. When subordinate females were removed from their groups and housed singly, but were maintained in scent contact with their dominant females, the onset of first ovulation was delayed to approximately 31 days in comparison to the average of 11 days in controls [15]. Visual cues from dominant females similarly delayed the onset of first ovulation in singly housed subordinate females rendered anosmic [14]. Intriguingly, the effectiveness of olfactory and visual cues to maintain ovarian inhibition lapsed after a few weeks. In addition, odor from a dominant female unfamiliar to the subordinate was ineffective in maintaining ovulatory inhibition in singly housed subordinates [10].

Although both olfactory and visual cues from dominant females may contribute to the maintenance of hypogonadotropism in subordinate female marmosets, behavioral cues from dominant females may provide the predominant inhibitory influence: subordinates rendered anosmic while remaining in their social groups failed to ovulate [14]. Together these findings suggest that subordinates may learn to associate olfactory and visual cues from an individual female with her dominance status. As a result, the dominant's olfactory and visual cues become conditioned stimuli contributing to hypogonadotropic anovulation in subordinates [10,89]. Associative learning may, therefore, figure prominently in the neural mechanisms translating social subordination into anovulation in female marmosets. This hypothesis, however, remains to be tested.

In addition to maintaining ovarian suppression, olfaction may play a predominant role in its initiation: five out of six female marmosets rendered anosmic prior to their introduction into a newly formed social group failed to demonstrate anovulation on becoming subordinate [5]. The mechanism of this important role of olfaction in the initiation of ovarian suppression is not yet clear. One possibility is that imprinting of olfactory cues from a dominant onto the neural substrate of subordinates might directly activate the neuroendocrine mechanisms inhibiting reproduction. Alternatively, olfactory cues from subordinates undergoing ovarian cycles might elicit aggressive behavior by the dominant, which in turn precipitates rapid inhibition of gonadotropin secretion and of ovulation [76].

A summary of our current understanding of the proximate cues and neuroendocrine mechanism(s) regulating inhibition of ovulation in subordinate female marmosets is shown in Fig. 4.



Fig. 5. (a) Antilog of the transformed mean (+95% confidence limit) of plasma concentrations of cortisol, DHEA and ACTH in 4–10 dominant female marmosets in the early to mid-follicular phase of the ovarian cycle and 4–14 anovulatory subordinate females. * P < 0.05 versus the dominant females, Student's *t*-test. (b) Antilog of the transformed mean (+95% confidence limit) of plasma concentrations of glucose following an overnight fast with or without a subsequent intravenous infusion of 120 mg of glucose in 4–6 dominant female marmosets in the early to mid-follicular phase of the ovarian cycle and in 4–6 anovulatory subordinate females. * P < 0.05 versus dominant females at 20 min and versus subordinate females at 0 min; Post-hoc testing following significant status × time interaction (P < 0.05) in a two-way ANOVA for repeated measures.

6. Adrenocortical and metabolic polyethism

Because subordinate female marmosets exhibited such pronounced reproductive polyethism, we examined two physiological components regulating homeostasis to determine if subordinate females demonstrated any metabolic accommodation of their hypoestrogenic infertile state.

6.1. Hypothalamic-pituitary-adrenal (HPA) axis

Within 5-8 weeks of becoming subordinate in a social group, female marmosets exhibit marked decreases in circulating cortisol concentrations (Fig. 5a; [10,76,79]). No such reductions in cortisol are noted in dominant females; thus, subordinate females in well established groups have significantly lower cortisol levels than dominants. This dichotomy in circulating cortisol concentrations was due to subordination per se and was not simply a consequence of decreased plasma estradiol levels secondary to ovulatory suppression in subordinate females: cortisol levels in hypoestrogenic, ovariectomized, pair-housed females did not fall to the same low level as in subordinates [10]. Circulating levels

of DHEA were also decreased in subordinate females (Fig. 5a). Taken together with the cortisol findings, this suggests that social subordination causes an inhibition of the steroidogenic activity of both the zona fasciculata and zona reticularis of the adrenal cortex in female marmosets.

The apparent reduction in steroidogenic output from the adrenal glands cannot be attributed to reduced pituitary secretion of ACTH, as circulating levels of ACTH do not differ between subordinate and dominant females (Fig. 5a; [78]). The reduced cortisol levels, however, may be due to decreased responsiveness of the adrenal cortex to ACTH stimulation, as dexamethasone-treated subordinate females showed poorer cortisol responses to an intravenous injection of ACTH₁₋₃₉ than similarly treated dominant females in the early to mid-follicular phase (Saltzman, W.; Schultz-Darken, N.J.; Abbott, D.H.; unpublished data).

Reduced HPA activity in subordinate female marmosets contrasts with findings from many competitively breeding species, in which subordinate individuals show elevated glucocorticoid levels as compared to dominants [10,76,79]. Interestingly, in several other cooperatively breeding species, reproductive suppression appears to be associated with either reduced glucorticoid levels, e.g. African wild dogs, *Lycaon pictus* [24]; dwarf mongoose, *Helogale parvula* [24]; Florida scrub jay, *Aphelocoma c. coerulescens* [85], or with glucocorticoid levels similar to those found in dominants, e.g. cotton-top tamarin, *Saguinus oedipus* [106]; black tufted-ear marmoset, *C. kuhli* [90]; Harris' hawk, *Parabuteo unicinctus* [63]. Thus, heightened HPA activity cannot be invoked as a cause of reproductive suppression in subordinate female marmosets and other cooperative breeders.

6.2. Glucose homeodynamics

Adrenal cortisol secretion in primates plays an important homeostatic role in regulating gluconeogenesis from the liver [68]. In human syndromes of chronically lowered adrenal cortisol secretion, such as Addison's disease, there are noticeable deficits in glucose homeodynamics [68]. Hypocortisolemic subordinate female marmosets, however, appear to be able to maintain euglycemia, since plasma glucose concentrations after an overnight fast were comparable to those of dominant females with normal cortisol levels (Fig. 5b). This short-term food deprivation tested the animals' ability to maintain normal circulating levels of glucose in the absence of sustenance. Nevertheless, when the marmosets were given an intravenous bolus of glucose following an overnight fast, to test their ability to take up and clear excess glucose from the circulation, subordinate females were slower in doing so than dominant females (Fig. 5b). The dominant females cleared excess glucose from their circulation within 20 min, whereas, the subordinates took 20-40 min to do so. This apparent difference between dominant and subordinate females in their ability to acutely regulate glucose uptake and clearance during an infusion of glucose suggests that compensatory metabolic processes [25] may not completely ameliorate the detrimental impact of chronic hypocortisolemia on glucose homeodynamics in subordinates.

7. Discussion

Subordinate female common marmosets exhibit a constellation of behavioral, neuroendocrinological and physiological differences from dominant females. The characteristics exhibited by subordinate female marmosets are strikingly similar to specializations made to accommodate subordinate status in social insects (Table 1) occupying comparable positions on the eusociality continuum proposed by Sherman and colleagues [86] (Fig. 1). According to the literature on invertebrate cooperative breeding, such adaptations to subordinate

status made by female marmosets should be considered as examples of polyethism, even if they are reversible by changes in social status [16]. In some of the social insects (e.g. paper wasps, Table 1), reversals in female status can occur naturally or can be hormonally induced experimentally. It is intriguing to speculate that specialized inhibitory neuroendocrine mechanisms are responsible for subordination-induced ovarian inhibition in these social insects, similar to our findings in common marmosets [10]. Such environmentally sensitive neuroendocrine mechanisms might be as labile as those found in female marmosets, to enable individuals to readily take advantage of appropriate changes in the social environment, such as the loss of the dominant breeding female, and permit previously subordinate females to effectively engage in competition with conspecifics for the vacant breeding position.

High degrees of intragroup relatedness may contribute significantly [42] to the extreme degrees of polyethism (phenotypically determined differences) and polymorphism (genetically determined differences) shown by some animals in their exquisite evolution of caste-structured eusocial societies (e.g. honey bees, yellow-jacket wasps, naked mole-rat (Heterocephalus glaber), Damaraland mole-rat (Cryptomys damarensis); Fig. 1). Little is known about intragroup relatedness in free-living groups of common marmosets, but an initial study of matrilines of mitochondrial D-loop DNA from groups at Nisia Floresta in northeastern Brazil suggests an average intragroup relatedness of less than 0.5 (C.G. Faulkes et al., personal communication). These molecular genetic studies of free-living marmosets also suggest that the breeding male in at least two social groups is genetically dissimilar to the other group members [35].

Since cooperatively breeding marmosets and some of the lower social insects appear to exhibit a number of analogous phenotypic and genotypic traits, such shared attributes may well reflect convergent adaptation to environmental conditions inauspicious for dispersal and independent breeding. Future comparative studies of behavioral, neuroendocrinological and physiological attributes of female subordinates in cooperatively breeding species may well reveal characteristics and mechanisms similar to those found in subordinate female marmosets, and would provide further support for considering cooperatively breeding vertebrates and invertebrates in an inclusive eusocial continuum [86]. In this regard, comparative neuroendocrinology provides a powerful tool in furthering our understanding of the proximate regulation of reproductive success in highly social species and may provide unique perspective and insight into both the neural mechanisms mediating environmental control of the hypothalamic-pituitarygonadal axis and the intrinsic factors shaping the evolution of sociality.

Table 1

Comparisons of behavioral, reproductive and non-reproductive polyethism characteristics of female common marmosets to those found in females in three groups of social insects exhibiting a similar reproductive skew to marmosets (Fig. 1)

Female characteristic	Marmosets ^a	Halictid bees ^b	Paper wasps ^c	Damp-wood termites ^d
Behavior				
A single, breeding female behaviorally dominates non-breeding (subor- dinate) females	Yes	Yes	Yes	Yes
Sexual behavior is more frequently exhibited by the dominant female	Yes	Yes ^e	Yes	Yes
Subordinate females preferentially forage in comparison to the domi- nant female	No	Yes	Yes	Yes
Subordinate females preferentially exhibit territorial defense in com- parison to the dominant female	Yes	Yes	Yes	Yes
Subordinate females play important roles in raising and foraging for the dominant female's offspring	Yes	Yes	Yes	Yes
Reproductive function				
Usually only one dominant female breeds	Yes	Yes ^e	Yes	Yes
The dominant female inhibits ovarian function in subordinates	Yes	Some	Yes	Yes
Specific neuroendocrine mechanisms (not stress) regulate ovarian inhi- bition in subordinates	Yes	?ť	Possibly ^g	?ť
Olfactory, visual and/or behavioral cues from the dominant female mediate ovarian inhibition in subordinates	Yes	Yes	Yes	Yes
Subordinate females can replace the dominant female as the breeding female, if the dominant is lost/removed	Yes ^h	Yes	Yes	Some ^g
Non-reproductive polyethism				
Dominant breeding females are larger then subordinate females	No	Yes	No ^j	Yes
Subordinate females form physically differentiated casts	No	No ^k	Slightly	Yes
Subordinate females are physiologically distinct from dominant fe- males	Yes	?ť	?t	?ť

^a This paper [40,97]; C. Lazaro-Parea and C.T. Snowdon, personal communication [10].

ь [16,65].

° [16,93].

^d [16,67].

^e Except for seasonal production of males [16].

^f Unknown.

^g Inhibitory neuroendocrine mechanisms have been demonstrated in subordinate females and are reversed by treatment with juvenile hormone or 20-hydroxyecdysone [72].

^h Except possibly in natal families which only contain 1⁰ relatives [77].

ⁱ Except for 'soldiers' [16].

^j Except in newly-established nests [16].

^k Except in very large colonies [16].

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References

- Abbott DH. Behavioral and physiological suppression of fertility in subordinate marmoset monkeys. Am J Primatol 1984;6:169–86.
- [2] Abbott DH. Natural suppression of fertility. In: Smith GR, Hearn JP, editors. Symposia of the Zoological Society of London, No. 60. New York: Oxford University Press, 1988:7– 28.
- [3] Abbott DH. Social conflict and reproductive suppression in marmosets and tamarin monkeys. In: Mason WA, Mendoza SP, editors. Primate Social Conflict. Albany: State University of New York Press, 1993:331–72.
- [4] Abbott DH. Social suppression of reproduction in primates. In: Standen V, Foley RA, editors. Comparative Socioecology. The Behavioural Ecology of Humans and other Mammals. Oxford: Blackwell, 1989:285–304.
- [5] Abbott DH, Faulkes CG, Barrett J, Smith TE, Cheesman DJ. Social control of female reproduction in marmoset monkeys and naked mole-rats. In: Lehnert H, Murison RH, Hellhammer

D, Beyer J, editors. Endocrine and Nutritional Control of Basic Biological Functions. Seattle: Hogrefe and Huber Publishers, 1993:475–90.

- [6] Abbott DH, George LM. Reproductive consequences of changing social status in female common marmosets. In: Box HO, editor. Primate Responses to Environmental Change. London: Chapman and Hall, 1991:294–309.
- [7] Abbott DH, George LM, Barrett J, Hodges KT, O'Byrne K, Sheffield JW, Sutherland IA, Chambers GR, Lunn SF, Ruiz de Elvira M-C. Social control of ovulation in marmoset monkeys: A neuroendocrine basis for the study of infertility. In: Ziegler TE, Bercovitch FB, editors. Socioendocrinology of Primate Reproduction. New York: Wiley-Liss, 1990:135–158.
- [8] Abbott DH, Hodges JK, George LM. Social status controls LH secretion and ovulation in female marmoset monkeys (*Callithrix jacchus*). J Endocrinol 1988;117:329–39.
- [9] Abbott DH, McNeilly AS, Lunn SF, Hulme MJ, Burden FJ. Inhibition of ovarian function in subordinate female marmoset monkeys (*Callithrix jacchus jacchus*). J Reprod Fertil 1981;63:335–45.
- [10] Abbott DH, Saltzman W, Schultz-Darken NJ, Smith TE. Specific neuroendocrine mechanisms not involving generalized stress mediate social regulation of female reproduction in cooperatively breeding marmoset monkeys. In: Carter CS, Lederhendler II, Kirkpatrick B, editors. Annal. N.Y. Acad. Sci, vol. 807. New York: New York Academy of Sciences, 1997:219–38.
- [11] Adler HJ, Jämmrich S. Breeding success in a polygynous group of *Callithrix jacchus* (L. 1758) in the allwetterzoo münster—a short report. Primate Rep 1991;31:105–7.
- [12] Alonso C. Failure to inhibit the reproduction of a subordinate female and change of hierarchy in a family group of *Callithrix jacchus jacchus*. A Primatol No Brasil 1986;2:203.
- [13] Anzenberger G. How stranger encounters of common marmosets (*Callithrix jacchus jacchus*) are influenced by family members: the quality of behavior. Folia Primatol 1985;45:204– 24.
- [14] Barrett J, Abbott DH, George LM. Sensory cues and suppression of reproduction in subordinate female marmoset monkeys, *Callithrix jacchus.* J Reprod Fertil 1993;97:301–10.
- [15] Barrett J, Abbott DH, George LM. Extension of reproductive suppression by pheromonal cues in subordinate female marmoset monkeys, *Callithrix jacchus*. J Reprod Fertil 1990;90:411–8.
- [16] Brian MV. Caste differentiation and division of labor. In: Herman HR, editor. Social Insects, vol. 1. New York: Academic Press, 1979:121–222.
- [17] Bronson FH. Mammalian Reproductive Biology. Chicago: University of Chicago Press, 1989.
- [18] Bronson FH, Heideman PD. Seasonal regulation of reproduction in mammals. In: Knobil E, Neill JD, editors. The Physiology of Reproduction, 2nd ed. New York: Raven Press, 1994:541-83.
- [19] Cardenas H, Ramirez VD. Maintenance of hypothalamic GnRH release during lactation in the rat: a push-perfusion study. Biol Res 1996;29:259–65.
- [20] Clarke IJ. Evidence that the switch from negative to positive feedback at the level of the pituitary gland is an important timing event for the onset of the preovulatory surge in LH in the ewe. J Endocrinol 1995;145:271–82.
- [21] Clarke IJ, Cummins JT. The temporal relationship between gonadotropin releasing hormone (GnRH) and luteinizing hormone (LH) secretion in ovariectomized ewes. Endocrinology 1982;111:1737–9.
- [22] Clutton-Brock TH, Albon SD, Guinness FE. Great expectations: dominance, breeding success and offspring sex ratios in red deer. Anim Behav 1986;34:460–71.

- [23] Cohen-Tannoudji J, Locatelli A, Signoret JP. Non-pheromonal stimulation by the male of LH release in the anoestrous ewe. Physiol Behav 1986;36:921–4.
- [24] Creel S, Creel NM, Monfort SL. Social stress in dominants. Nature 1996;379:212.
- [25] Cryer PE. Glucose homeostasis and hypoglycemia. In: Wilson JD, Foster DW, editors. Williams Textbook of Endocrinology, 8th ed. Philadelphia, PA: Saunders, 1992:1223–53.
- [26] de Catanzaro D, MacNiven E. Psychogenic pregnancy disruptions in mammals. Neurosci Biobehav Rev 1992;16:43–53.
- [27] Digby LJ. Infant care, infanticide and female reproductive strategies in polygynous groups of common marmosets (*Callithrix jacchus*). Behav Ecol Sociobiol 1995;37:51–61.
- [28] Digby LJ, Ferrari SF. Multiple breeding females in free-ranging groups of *Callithrix jacchus*. Int J Primatol 1994;15:389–97.
- [29] Emlen ST. The evolution of cooperative breeding in birds and mammals. In: Krebs JR, Davies NB, editors. Behavioural Ecology: An Evolutionary Approach, 3rd ed. Oxford: Blackwell, 1991:301–37.
- [30] Epple G. Vergleichende Untersuchungen über Secual- und Sozialverhalten der Krallenaffen (Hapalidae). Folia Primatol 1967;8:1–40.
- [31] Epple G, Belcher AM, Kuederling I, Zeller U, Scolnick L, Greenfield KL, Smith AB III. Making sense out of scents: species differences in scent glands, scent-marking behaviour and scent-mark composition in the Callitrichidae. In: Rylands AB, editor. Marmosets and Tamarins: Systematics, Behaviour and Ecology. London: Oxford University Press, 1993:123–51.
- [32] Estep DQ, Nieuwenhuijsen K, Bruce KEM, de Neef KJ, Walters PA III, Baker SC, Slob AK. Inhibition of sexual behaviour among subordinate stumptail macaques, *Macaca arctoides*. Anim Behav 1988;36:854–64.
- [33] Evans JJ. Oxytocin and the control of LH. J Endocrinol 1996;151:169-74.
- [34] Faulkes CG, Abbott DH. The physiology of a reproductive dictatorship: regulation of male and female reproduction by a single breeding female in colonies of naked mole-rats. In: Solomon NG, French JA, editors. Cooperative Breeding in Mammals. Cambridge: Cambridge University Press, 1997:302– 34.
- [35] Faulkes CG, Arruda MF, Monteiro da Cruz MAO. Mitochondrial DNA control region sequence divergence within and between wild populations of the common marmoset, *Callithrix jacchus*, IPS/APS Cong Abs 1996:781.
- [36] Ferin M, vanVught D, Wardlaw S. The hypothalamic control of the menstrual cycle and the role of endogenous opioid peptides. Rec Prog Horm Res 1984;40:441–85.
- [37] Ferrari SF, Diego VH. Long-term changes in a wild marmoset group. Folia Primatol 1992;58:215–8.
- [38] Ferrari SF, Lopes Ferrari MA. A re-evaluation of the social organization of the Callitrichidae, with reference to the ecological differences between genera. Folia Primatol 1989;52:132–47.
- [39] Ferrari SF, Digby LJ. Wild *Callithrix* groups: stable extended families? Am J Primatol 1996;38:19–27.
- [40] French JA. Proximate regulation of singular breeding in Callitrichid primates. In: Solomon NG, French JA, editors. Cooperative Breeding in Mammals. Cambridge: Cambridge University Press, 1997:34–75.
- [41] Haigh G, Cushing BS, Bronson FH. A novel post-copulatory block of reproduction in white-footed mice. Biol Reprod 1988;38:623-6.
- [42] Hamilton WD. The genetical evolution of social behavior, I, II. J Theor Biol 1964;7:1–52.
- [43] Harlow CR, Hillier SG, Hodges JK. Androgen modulation of follicle-stimulating hormone-induced granulosa cell steroidogenesis in the primate ovary. Endocrinology 1986;119:1403–5.

- [44] Harlow CR, Shaw HJ, Hillier SG, Hodges JK. Factors influencing follicle-stimulating hormone-responsive steroidogenesis in marmoset granulosa cells: effects of androgens and the stage of follicular maturity. Endocrinology 1988;122:2780–7.
- [45] Hearn JP. Restraining device for small monkeys. Lab Anim 1977;11:261–1.
- [46] Hillier SG, Harlow CR, Shaw HJ, Wickings EJ, Dixson AF, Hodges JK. Granulosa cell differentiation in primate ovaries: the marmoset monkey (*Callithrix jacchus*) as a laboratory model. In: Stouffer RL, editor. The Primate Ovary. New York: Plenum, 1987:61–73.
- [47] Hillier SG, Harlow CR, Shaw HJ, Wickings EJ, Dixson AF, Hodges JK. Cellular aspects of pre-ovulatory folliculogenesis in primate ovaries. Hum Reprod 1988;3:507–11.
- [48] Hubrecht RC. Field observations on group size and composition of the common marmoset (*Callithrix jacchus jacchus*), at Tapacura, Brazil. Primates 1984;25:13–21.
- [49] Hubrecht RC. The fertility of daughters in common marmoset (*Callithrix jacchus jacchus*) family groups. Primates 1989;30:423–32.
- [50] Huck UW, Bracken AC, Lisk RD. Female induced pregnancy block in the golden hamster. Behav Neurol Biol 1983;38:190–3.
- [51] Hunter AJ, Fleming D, Dixson AF. The structure of the vomeronasal organ and nasopalatine ducts in *Aotus trivirgatus* and some other primate species. J Anat 1984;138:217–25.
- [52] Jennions MD, MacDonald DW. Cooperative breeding in mammals. Trends Ecol Evol 1994;9:89–93.
- [53] Kaplan JR, Adams MR, Koritnik DR, Rose JC, Manuck SB. Adrenal responsiveness and social status in intact and ovariectomized *Macaca fascicularis*. Am J Primatol 1986;11:181–93.
- [54] Karsch FJ, Dahl GE, Evans NP, Manning JM, Mayfield KP, Moenter SM, Foster DL. Seasonal changes in gonadotropin-releasing hormone secretion in the ewe: alteration in response to the negative feedback action of estradiol. Biol Reprod 1993;49:1377–83.
- [55] Keller L, Perrin N. Quantifying the level of eusociality. Proc R Soc London B 1995;260:311–5.
- [56] Koenig A, Rothe H. Social relationships and individual contribution to cooperative behaviour in captive common marmosets (*Callithrix jacchus*). Primates 1991;32:183–95.
- [57] Lacey EA, Sherman PW. Cooperative breeding in naked molerats: implications for vertebrate and invertebrate sociality. In: Solomon NG, French JA, editors. Cooperative Breeding in Mammals. Cambridge: Cambridge University Press, 1997:267– 301.
- [58] Levine JE, Kwok-Yuen FP, Ramirez VD, Jackson GL. Simultaneous measurement of luteinizing hormone-releasing hormone and luteinizing hormone release in unanesthetized, ovariectomized sheep. Endocrinology 1982;111:1449–55.
- [59] Lincoln GA. Photo-neuroendocrine control of seasonal cycles in body weight, pelage growth and reproduction in rams. This volume, 1997.
- [60] Maeda K-I, Tsukamara H, Uchida E, Okhura N, Okhura S, Yoloyama A. Changes in the pulsatile secretion of LH after the removal of and subsequent resuckling by pups in ovariectomized lactating rats. J Endocrinol 1989;121:277–83.
- [61] Martin RD, Dixson AF, Wickings EJ, editors. Paternity in Primates: Genetic Tests and Theories. Basel: Karger, 1992.
- [62] Martin GB, Oldham CM, Lindsay PR. Increased plasma LH levels in seasonally anovular merino ewes following the introduction of the rams. Anim Reprod Sci 1980;3:125–32.
- [63] Mays NA, Vleck CM, Dawson J. Plasma luteinizing hormone, steroid hormones, behavioral role and nest stage in cooperatively breeding Harris' hawks (*Parabuteo unicinctus*). Auk 1991;108:619–37.
- [64] Mendes Pontes AR, Monteiro da Cruz MAO. Home range, intergroup transfers and reproductive status of common mar-

mosets *Callithrix jacchus* in a forest fragment in north-eastern Brazil. Primates 1995;36:335–47.

- [65] Michener CD. Comparative social behavior of bees. Ann Rev Entomol 1969;14:299–342.
- [66] Moenter SM, Caraty A, Karsch FJ. The estradiol-induced surge of gonadotropin-releasing hormone in the ewe. Endocrinology 1990;127:1375–84.
- [67] Noirot C. Sexual castes and reproductive strategies in termites. In: Engels W, editor. Social Insects. An Evolutionary Approach to Castes and Reproduction. Berlin: Springer-Verlag, 1990:5– 35.
- [68] Orth DN, Kovacs WJ, DeBold CR. The adrenal cortex. In: Wilson JD, Foster DW, editors. Williams Textbook of Endocrinology, 8th ed. Philadelphia, PA: Saunders, 1992:489–619.
- [69] Pemberton JM, Albon SD, Guiness FE, Clutton-Brock TH, Denver GA. Behavioral estimates of male mating success tested by DNA fingerprinting. In: Krebs JR, Davies NB, editors. Behavioral Ecology, vol 3. Oxford, UK: Blackwell Scientific Publications, 1992:66–75.
- [70] Rhine RJ, Wasser S, Norton GW. Eight-year study of social and ecological correlates of mortality among immature baboons of Mikumi National Park, Tanzania. Am J Primatol 1988;16:199–212.
- [71] Roda SA, Roda S. Infanticide in a natural group of *Callithrix jacchus* (Callithrichidae-Primates). Int J Primatol 1987;8:497.
- [72] Roseler PF. Endocrine basis of dominance and reproduction in polistine paper wasps. Fortschr Zool 1985;31:259–72.
- [73] Rothe H. Some aspects of sexuality and reproduction in groups of captive marmosets (*Callithrix jacchus*). Z Tierpsychol 1975;37:255–73.
- [74] Rothe H, Koenig A. Variability of social organization in captive common marmosets (*Callithrix jacchus*). Folia Primatol 1991;57:28–33.
- [75] Rylands AB. Habitat and the evolution of social and reproductive behavior in Callitrichidae. Am J Primatol 1996;38:5–18.
- [76] Saltzman W, Schultz-Darken NJ, Abbott DH. Behavioural and endocrine predictors of dominance and tolerance in female common marmosets, *Callithrix jacchus*. Anim Behav 1996;51:657–74.
- [77] Saltzman W, Schultz-Darken NJ, Abbott DH. Familial influences on ovulatory function in common marmosets (*Callithrix jacchus*). Am J Primatol 1997;41:159–77.
- [78] Saltzman W, Schultz-Darken NJ, Abbott DH. Social and reproductive influences on plasma cortisol and ACTH levels in female common marmosets: sensitivity to glucocorticoid negative feedback. Am J Primatol 1997;42:144–5.
- [79] Saltzman W, Schultz-Darken NJ, Scheffler G, Wegner FW, Abbott DH. Social and reproductive influences on plasma cortisol in female marmoset monkeys. Physiol Behav 1994;45:801–10.
- [80] Saltzman W, Schultz-Darken NJ, Severin JM, Abbott DH. Escape from social suppression of sexual behavior and of ovulation in female common marmosets. In: Carter CS, Lederhendler II, Kirkpatrick B, editors. Ann. New York Acad. Sci, vol. 807. New York: New York Academy of Sciences, 1997:567–70.
- [81] Saltzman W, Schultz-Darken NJ, Terasawa E, Abbott DH. In vivo release of gonadotropin-releasing hormone (GnRH) in socially subordinate, anovulatory female marmoset monkeys. Soc Neurosci Abstr 1995;21:265.
- [82] Saltzman W, Severin JM, Schultz-Darken NJ, Abbott DH. Behavioral and social correlates of escape from suppression of ovulation in female common marmosets housed with the natal family. Am J Primatol 1997;41:1–21.
- [83] Savage A, Giraldo LH, Soto LH, Snowdon CT. Demography, group composition and dispersal in wild cotton-top tamarin (*Saguinus oedipus*) groups. Am J Primatol 1996;38:85–100.

- [84] Scanlon CE, Chalmers NR, Monteiro da Cruz MAO. Changes in the size, composition and reproductive condition of wild marmoset groups (*Callithrix jacchus jacchus*) in north east Brazil. Primates 1988;29:295–305.
- [85] Schoech SJ, Mumme RL, Moore MC. Reproductive endocrinology and mechanisms of breeding inhibition in cooperatively breeding Florida scrub jays (*Aphelocoma c. coerulescens*). Condor 1991;93:354–64.
- [86] Sherman PW, Lacey EA, Reeve HK, Keller L. The eusociality continuum. Behav Ecol 1995;6:102–8.
- [87] Signoret JP, Lindsay DR. The male effect in domestic animals: effect on LH secretion and ovulation. Importance of olfactory cues. In: Briephol W, editor. Olfaction and Endocrine Regulation. London: IRL Press, 1982:63–72.
- [88] Smith MS, Neill JD. Inhibition of gonadotropin secretion during lactation in the rat: relative contribution of sucking and ovarian steroids. Biol Reprod 1977;17:255–61.
- [89] Smith TE, Abbott DH, Tomlinson AJ, Mlotkiewicz JA. Differential display of investigative behavior permits discrimination of scent signatures from familiar and unfamiliar socially dominant female marmoset monkeys (*Callithrix jacchus*), J Chem Ecol 1997;23:2523–46.
- [90] Smith TE, French JA. Social and reproductive conditions modulate urinary cortisol excretion in black tufted-ear marmosets (*Callithrix khuli*). Am J Primatol 1997;42:253–67.
- [91] Solomon NG, French JA. Cooperative Breeding in Mammals. Cambridge: Cambridge University Press, 1997.
- [92] Stevenson MF, Rylands AB. The marmosets, genus *Callithrix*. In: Mittermeier RA, Rylands AB, Coimbra-Filho AF, da Fonseca GAB, editors. Ecology and Behavior of Neotropical Primates, vol. 2. Washington, DC: World Wildlife Fund, 1988:131–222.
- [93] Strambi A. Physiology and reproduction in social wasps. In: Engels W, editor. Social Insects. An Evolutionary Approach to Castes and Reproduction. Berlin: Springer-Verlag, 1990:59–75.
- [94] Summers PM, Wennick CJ, Hodges JK. Cloprostenol-induced luteolysis in the marmoset monkey (*Callithrix jacchus*). J Reprod Fertil 1985;73:133–8.
- [95] Sussman RW, Garber PA. A new interpretation of the social organization and mating system of Callitrichidae. Int J Primatol 1987;8:73–92.

- [96] Taniguchi K, Matsusaki Y, Ogawa K, Saito TR. Fine structure of the vomernasal organ in the common marmoset (*Callithrix jacchus*). Folia Primatol 1992;59:169–76.
- [97] Tardif SD. The bioenergetics of parental behavior and the evolution of alloparental care in marmosets and tamarins. In: Solomon NG, French JA, editors. Cooperative Breeding in Mammals. Cambridge: Cambridge University Press, 1997:11– 33.
- [98] Terasawa E, Gearing M. Luteinizing hormone releasing hormone (LHRH) neuroterminals mapped using the push-pull perfusion method in the rhesus monkey. Brain Res Bull 1988;21:117–21.
- [99] Terasawa E, Krook C, Hei DL, Gearing M, Schultz NJ, Davis GA. Norepinephrine is a possible neurotransmitter stimulating pulsatile release of luteinizing hormone-releasing hormone in the rhesus monkey. Endocrinology 1988;123:1808–16.
- [100] Torii R, Abbott DH, Nigi H. Morphological changes of the ovary and hormonal changes through the ovarian cycle of the common marmoset (*Callithrix jacchus*). Primates 1996;37:49– 56.
- [101] Wasser SK, Barash DP. Reproductive suppression among female mammals: Implications for biomedicine and sexual selection theory. Q Rev Biol 1983;58:513–38.
- [102] Watanabe G, Terasawa E. In vivo release of luteinizing hormone releasing hormone increases with puberty in the female rhesus monkey. Endocrinology 1989;125:92–9.
- [103] Wilson EO. Sociobiology. Boston: Harvard University Press, 1975.
- [104] Yasin M, Dalkin AC, Haisenleder DD, Marshall JC. Testosterone is required for gonadotropin-releasing hormone stimulation of luteinizing hormone- β messenger ribonucleic acid expression in female rats. Endocrinology 1997;137:1265–71.
- [105] Zanuy S, Carrillo M, Mateos J, Trudeau V, Hah O. The effects of sustained administration of testosterone on brain and pituitary sGnRH levels, plasma sex steroids and gonadal development in immature sea bass, *Dicentrarchus labrax*. This volume, 1997.
- [106] Ziegler TE, Scheffler G, Snowdon CT. The relationship of cortisol levels to social environment and reproductive functioning in female cotton-top tamarins, *Saguinus oedipus*. Horm Behav 1995;29:407–24.